

# CELL DATA BASE

#### PREPARED BY

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### **CELL EXPANSION**

Cell culture is an essential procedure in life sciences where all cell types need to be maintained under lab conditions.

Human Platelet Lysate (HPL) resembles the natural cell environment for optimal growth results.

This Cell Database is based on customer feedback, publications and internal tests and investigations – summarizing cells which have shown favourable properties grown in Human Platelet Lysate.



01

### HUMAN PRIMARY CELLS

List of human primary cells that showed favourable properties with HPL.

02

### HUMAN CELLS LINES

List of human cell lines that showed favourable properties with HPL.

03

#### ANIMAL PRIMARY CELLS

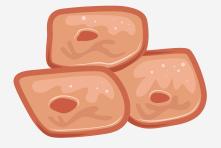
List of animal primary cells that showed favourable properties with HPL.

ANIMAL CELL LINES

List of animal cell lines that showed favourable properties with HPL.



# HUMAN PRIMARY CELLS

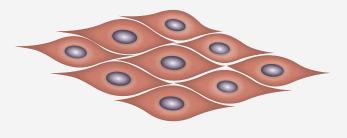


| Cell Type                                   | Short Form |
|---|------------|
| Adipose-derived Stem Cells                  | ADSC       |
| Chondrocytes                                |            |
| Corneal keratocytes                         |            |
| Dental Follicular Cells DFSC                |            |
| Fibroblasts                                 |            |
| Dermal Fibroblasts                          |            |
| Foreskin Fibroblasts                        |            |
| Gamma-Delta-T-Cells                         |            |
| Head and Neck Squamous Cell Carcinoma Cells | PCI 13     |
| Hematopoietic Stem Cells                    | HSCs       |
| Umbilical vein endothelial cells            | HUVEC      |
| Neural crest cells                          |            |
| Lymphocytes from Blood                      |            |
| Macrophages/Monocytes                       |            |
| Peridontal Ligaments Cells                  |            |
| Neural crest cells                          | PDL        |
| Stem Cells from Sweat Glands                | SGSCs      |
| Human Dental Pulp Stem cells                | DPSC       |

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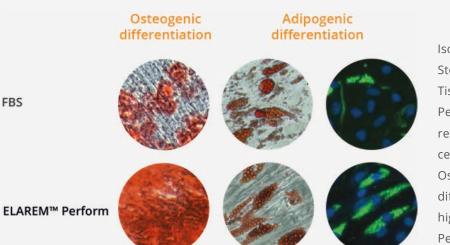
# HUMAN PRIMARY CELLS



### Mesenchymal Stromal Cells (MSCs)

We dedicate a full page to these special cells, also known as Mesenchymal Stem Cells. Due to their multipotency, they are able to differentiate into a variety of cell types. This characteristic makes them interesting cells used in cell culture research and therapy.

| Cell Type  | Short Form |
|--|------------|
| Mesenchymal Stem Cells from adipose tissue       | MSC-AT     |
| Mesenchymal Stem Cells from bone marrow          | MSC-BM     |
| Mesenchymal Stem Cells from MNCs                 | MSC-MNC    |
| Mesenchymal Stem Cells from umbilical cord       | MSC-UC     |
| Mesenchymal Stem Cells differentiated from iPSCs | iPS-MSCs   |



Isolation of Mesenchymal Stem Cells from Adipose Tissue with 10% ELAREM™ Perform or 10% FBS resulted in MSCs of similar cellular morphology. Osteogenic and adipogenic differentiation of MSCs was higher with ELAREM™ Perform than with FBS.



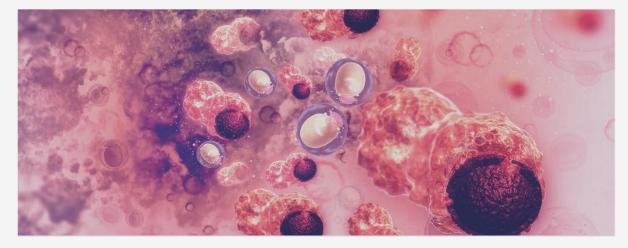
# HUMAN CELL LINES



Short Form

## Cell Type

| Breast Carcinoma                           | BT-20  |
|--|--------|
| Breast Carcinoma                           | HBL100 |
| Cervical cancer cells                      | HeLa   |
| Colon cancer cell lines                    | LS 180 |
| Epithelial colorectal adenocarcinoma cells | Caco-2 |
| Lung Large-cell carcinoma LCC              |        |
| Melanoma                                   |        |
| Osteo Sarcoma HOS(TE85)                    |        |
| Human osteosarcoma cell line               | U-2 OS |
| Human urinary bladder carcinoma cell line  | 5637   |
| Human lung adenocarcinoma cell line        | HGC-27 |



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# HUMAN CELL LINES



Short Form

## Cell Type

| Chordoma cell lines                             |           |
|---|-----------|
| Dermal Fibroblasts                              | HFFF2     |
| Dermal keratinocytes                            | NCTC 2544 |
| Epithelia mammary gland; breast/duct            | ZR-75-1   |
| Human embryonal lung fibroblast                 | MRC-5     |
| Human Embryonic Kidney 293 cells                | HEK-293   |
| Human Gingiva Fibroblasts                       | HGF-1     |
| Human umbilical vein endothelial cells          | HUVEC     |
| Keratinocyte cell line from adult human skin    | НаСаТ     |
| Hematopoietic Stem Cells                        | HSCs      |
| Lymphocytes (immortalized)                      |           |
| MSCs containing catalytic subunit of telomerase | hMSC-TERT |
| Retinal pigmented epithelium                    | ARPE-19   |
| Human epithelial type 2                         | HEp-2     |
| Human colorectal adenocarcinoma cell line       | HROC24    |
| Pancreas adenocarcinoma cell line               | Panc-1    |
| Human renal clear cell carcinoma cell line      | RCC-ER    |

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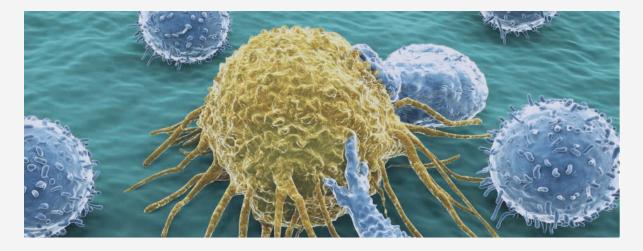
# HUMAN CELL LINES



Short Form

## Cell Type

| Human continous tumor                    | A-549    |
|--|----------|
| Human continous tumor                    | Caco-2   |
| Human continous tumor                    | MCF-7    |
| Human continous tumor                    | U-251 MG |
| Human leukemia cells                     | HL-60    |
| Human leukemia cells                     | Jurkat   |
| Human leukemia cells                     | KG-1     |
| Myelogenous leukemia                     | K562     |
| Human myeloid leukemia cell line         | Kasumi-1 |
| Human acute monocytic leukemia cell line | THP-1    |

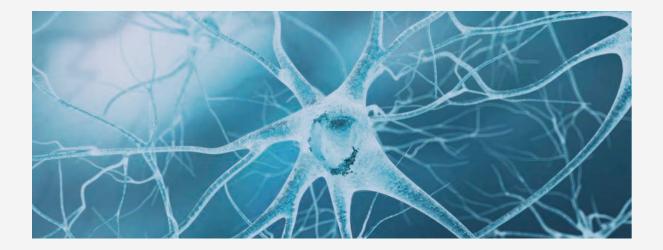




# ANIMAL PRIMARY CELLS



| Cell Type                                 | Short Form |  |
|---|------------|--|
| Bovine Corneal Endothelial Cells          | CEC        |  |
| Murine Astrocytes                         |            |  |
| Murine Mesenchymal Stem Cells             | MSC        |  |
| Rat Mesenchymal Stem Cells                | MSC        |  |
| Murine Mikroglia                          |            |  |
| Spiral ganglions from Sprague-Dawley rats | SGC        |  |





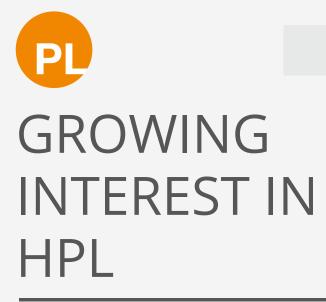
# ANIMAL CELL LINES

## Cell Type

| Adrenal gland                        | PC-12     |
|--------------------------------------|-----------|
| Aurenai gianu                        | PC-12     |
| African Green Monkey Fibroblast      | COS-7     |
| Chinese Hamster Ovary epithelial     | СНО       |
| Kidney (African Green Monkey)        | Vero      |
| Mouse adenocarcinoma cell line       | RAG       |
| Mouse fibroblast cell line L929      |           |
| Mouse mammary tumor 060562 MMT 06056 |           |
| Mouse Mikoglia BV-2                  |           |
| Mouse myeloma cell line              | Sp2O-Ag14 |
| Neuroblastoma cell line              | Neuro-2a  |
| Statens Seruminstitut Rabbit Cornea  | SIRC      |
| Testis from rat                      | R2C       |

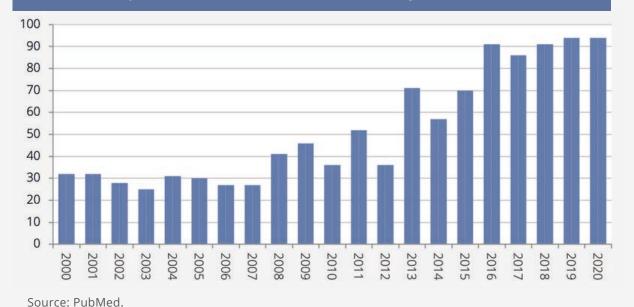
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Short Form





### Number of publications on HPL over the years



### Interest in Human Platelet Lysate emerges

The number of Human Platelet Lysate publications has been increasing steadily for years. This also results in a growing number of tested cells of <u>different origins.</u> PL BioScience contributes to this trend by being part of several research projects as well as performing own laboratory tests.

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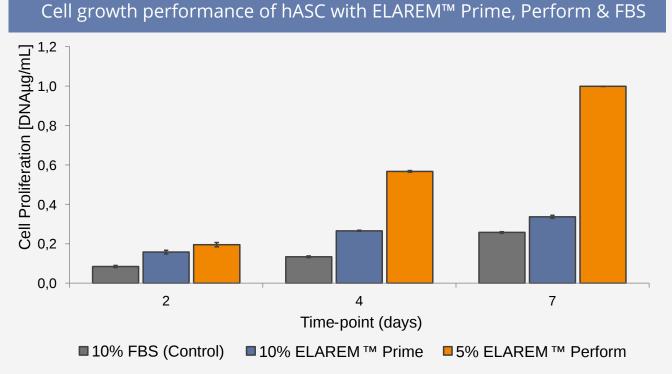


## PERFORMANCE

Human Platelet Lysate contains abundant growth factors and cytokines derived from human platelets. Those factors have been shown to stimulate cellular proliferation and maintain phenotype and differentiation potential of various cells.

Final concentration of ELAREM<sup>™</sup> human Platelet Lysate in cell culture medium can vary depending on cell type and experimental conditions. It is recommended to determine the optimal concentration – between 1% and 10% (v/v) – for the cells of interest.

"HPL can support the growth and proliferation of cells in culture, as platelets are known to play a vital role in tissue renewal and wound healing." (1)



(1) Human Platelet Lysate efficiency, stability, and optimal heparin concentration required in culture of mammalian cells. Mohamed et al. (2020), The Korean Journal of Hematology. Blood Res 2020; 55(1): 35-43, Internet: https://www.bloodresearch.or.kr/journal/view.html? doi=10.5045/br.2020.55.1.35

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## OUR COMPANY'S VISION FOR THIS PROJECT

We at PL BioScience do our best to enhance the advances in regenerative medicine with our animal-free cell culture tools. As a result, we produce Human Platelet Lysates which bear the potential to raise both stem cell research and therapy to the next level.

Our aim and our vision: The future of cell culture supplements is animal-free.

To achieve this aim,

- we raise awareness of alternatives to animal sera
- share scientific knowledge about Human Platelet Lysate and
- collect data about cells, cell lines and innovations in cell culture.

You can help us to achieve this aim! Share your experiences and knowledge about cells and HPL with us.



## MEET OUR SUPPORT TEAM



support@pl-bioscience.com



#### **DR. SILKE ISENHARDT**

Product Support Specialist

As a biologist, Silke is our expert for cells and has deep knowledge about Human Platelet Lysate. She is your contact person when you have a question about our products and their handling.



### PHILIPP SCHMIDT

Sales Specialist

A sales expert with long experience in B2B sales and customer support. Philipp is your contact person when you have a question about products, prices or contracts.



### SONJA ZADEL

Internal Sales Specialist

Sonja is our expert for internal sales coordination including import and export. She is your contact person when you have a question about your delivery or product

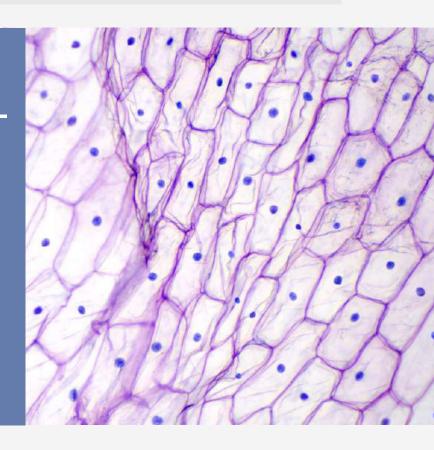


### SOURCES

This Cell Database is based on customer feedback, publications and internal tests and investigations.

We constantly strive to expand our knowledge and by this, our cell database.

If you plan a project with cells that are not included in this list, please contact us.



## OWN PUBLICATIONS

| Epigenetic Biomarker to Support Classification into Pluripotent and Non-Pluripotent Cells.<br>Lenz M., Goetzke R., Schenk A., Schubert C., Veeck J., Hemeda H., Koschmieder S., Zenke M.,<br>Schuppert A. & Wagner W. (2015). Scientific Reports; 5:8973. |
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| Evaluation of human platelet lysate versus fetal bovine serum for culture of mesenchymal stromal cells.   |
| Hemeda H., Wagner W. & Giebel B. (2014). Cytotherapy; 16(2):170-180.  |
| Epigenetic Rejuvenation of Mesenchymal Stromal Cells Derived from Induced Pluripotent<br>Stem Cells.<br>Frobel F., Hemeda H., Lenz M., Abagnale G., Joussen S., Denecke B., Šarić T., Zenke M. &<br>Wagner W. (2014). Stem Cell Reports; Vol. 3; 414–422. |
| Matrix elasticity, replicative senescenece and DNA methylation patterns of mesenchymal stem cells.  |
| Schellenberg A., Joussen S., Moser K., Hampe N., Hersche N., Hemeda H., Schnitker J.,   |
| Denecke B., Qiong L., Pallua N., Zenke M., Merkel R., Hoffmann B. & Wagner W. (2014).   |
| Biomaterials; 35(24):6351-6358.   |





## OWN PUBLICATIONS

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Donor age of human platelet lysate affects proliferation and differentation of mesenchymal stem cells.

Lohmann M., Walenda G., Hemeda H., Joussen S., Drescher W., Jockenhoevel S., Hutschenreuter G., Zenke M. & Wagner W. (2012). PLoS ONE; 7(5): e37839.

Human platelet lysate gel provides a novel three dimensional-matrix for enhanced culture expansion of mesenchymal stromal cells.

Walenda G., Hemeda H., Schneider R. K., Merkel R., Hoffmann B. & Wagner W. (2012). Tissue Eng Part C Methods; 18(12):924-934.



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| A High-Throughput Method as a Diagnostic Tool for HIV Detection in Patient-Specific Induced<br>Pluripotent Stem Cells Generated by Different Reprogramming Methods.<br>Hübscher D, Rebs S, Haupt L, et al. (2019). Stem Cells International.  |
| An injectable platelet lysate-hyaluronic acid hydrogel supports cellular activities and induces<br>chondrogenesis of encapsulated mesenchymal stem cells.<br>Jooybar E, Abdekhodaie MJ, Alvi M, et al. (2019). Acta Biomaterialia; 83: 233–244.   |
| CD10/Neprilysin Enrichment in Infrapatellar Fat Pad-Derived Mesenchymal Stem Cells Under<br>Regulatory-Compliant Conditions: Implications for Efficient Synovitis and Fat Pad Fibrosis<br>Reversal.<br>Kouroupis D, Bowles A, Best T, et al. (2020). The American Journal of Sports Medicine, 1-15. |
| Development and Characterization of a Parallelizable Perfusion Bioreactor for 3D Cell<br>Culture.<br>Egger D, Fischer M, Clementi A, et al. (2017). Bioengineering; 4:51.   |
| Fetal Bovine Serum (FBS): Past – Present – Future. Consensus Report.<br>van der Valk J, Bieback K, Buta C, et al. (2018). ALTEX – Alternatives to animal<br>experimentation; 35(1):99-118.  |



## PRODUCT REFERENCES

| From 3D to 3D: isolation of mesenchymal stem/stromal cells into a three-dimensional<br>human platelet lysate matrix.<br>Egger D, Oliveira A, Mallinger B, et al. (2019). Stem Cell Research & Therapy; 10:248.   |
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| Human Platelet Lysate Can Replace Fetal Calf Serum as a Protein Source to Promote<br>Expansion and Osteogenic Differentiation of Human Bone-Marrow-Derived Mesenchymal<br>Stromal Cells.<br>Karadjian M, Senger A-S, Essers C, et al. (2020). Cells; 9: 918. |
| Human platelet lysate current standards and future developments. Committee report.<br>Henschler R, Gabriel C, Schallmoser K, et al. (2019). Transfusion; 9999; 1–7.  |
| Hypoxia Conditioned Mesenchymal Stem Cell-Derived Extracellular Vesicles Induce Increased<br>Vascular Tube Formation in vitro.<br>Almeria C, Weiss R, Roy M, et al. (2019). Front. Bioeng. Biotechnol. 7:292.  |
| Hypoxic Three-Dimensional Scaffold-Free Aggregate Cultivation of Mesenchymal Stem Cells<br>in a Stirred Tank Reactor.<br>Egger D, Schwedhelm I, Hansmann J, et al. (2017). Bioengineering; 4, 47.  |
| Platelet lysate outperforms FCS and human serum for co-culture of primary human<br>macrophages and hMSCs.<br>Tylek T, Schilling T, Schlegelmilch K, et al. (2019). Scientific Reports; 9:3533.   |



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Pluripotent Stem Cell-Derived Mesenchymal Stem Cells Show Comparable Functionality to Their Autologous Origin.

Jakob M, Hambrecht M, Spiegel J, et al. (2020). Cells; 10:33.

Regulatory-compliant conditions during cell product manufacturing enhance in vitro immunomodulatory properties of infrapatellar fat pad-derived mesenchymal stem/stromal cells.

Kouroupis D, Bowles A, Greif D, et al. (2020). Cytotherapy.

The Power of LC-MS Based Multiomics: Exploring Adipogenic Differentiation of Human Mesenchymal Stem/Stromal Cells. Rampler E, Egger D, Schoeny H, et al. (2019). Molecules; 24:3615.



## GENERAL PUBLICATIONS

Culture of human cell lines by pathogen-inactivated human Platelet Lysate Fazzina R, Iduicone P, Mariotti A, Fioravanti D, et al. (2015). Springer; DOI 10.1007/s10616-015-9878-5

Differentiation of Rat bone marrow Mesenchymal stem cells into Adipocytes and Cardiomyocytes after treatment with Platelet Lysate. Moghadam F H, Tayeb T, Barzegar K, et al. (2015). IJOSCR, Volume 10, Number 1

Effect of Platelet Lysate on Human Cells Involved in Different Phases of Wound Healing. Barsotti M C, Losi P, Briganti E, Sanguinetti E et al. (2013). PLOS ONE, Volume 8, Issue 12, e84753

Ex vivo Expansion of Bovine Corneal Endothelial Cells in Reno-Free Medium Supplemented with Platelet Releasate Chou M L, Burnouf T, Wang TJ, et al. (2014). PLOS ONE, Volume 9, Issue 6, e99145

Human Platelet Lysate efficiency, stability, and optimal heparin concentration required in culture of mammalian cells. Mohamed et al. (2020), The Korean Journal of Hematology. Blood Res 2020; 55(1): 35-43, Internet: https://www.bloodresearch.or.kr/journal/view.html? doi=10.5045/br.2020.55.1.35

Platelet Lysate Inhibits NF-kB Activation and Induces Proliferation and an Alert State in Quiescent Human Umbilical Vein Endothelial Cells Retaining Their Differentiation Capability. Romaldini A, Ulivi V, Nardini M, et al. (2018). Cells; 8, 331; doi:10.3390/cells8040331

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